DNA SEQUENCING ORDER SHEET (rev 02/21/03)D.B.

Name: _____ Principle Investigator: _____
Phone # _____ Date: _____
Room # _____ Email address:
_____

Template ID (e.g. JS21)	**Template Concentration** (100 ng/µl Plasmid) (20 ng/µl PCR)**	**Primer Name**	**Primer Concen-tration (need 5-10 µM)**	**Template Size (kB)**	**Notes (see below)** **If PCR product or BAC please note that in this column**

INSTRUCTIONS:

Template ID: use the laboratory P.I.'s initials and then number each DNA sample sequentially with every submission, e.g. JS10, JS11, JS12 etc. The template ID should also be used to label the tube of DNA submitted, **on the top of each tube**. If the same DNA is being tested with more than one primer, you may put the DNA in a single tube, but it must be labeled with more than one Template ID number, e.g. JS10-13

****Template concentration:** SAMPLES MUST BE IN WATER. **For PLASMID and BAC samples, the DNA concentration should be 100ng/µl -** please submit **800ng** (=8 µl at 100ng/µl) of DNA per sequencing **reaction**. **For PCR products**, the DNA concentration should be 20 ng/µl, and you need to submit **10ng PCR product DNA/100bases length**. (Please indicate SIZE of the PCR product in the *Template size* column above; also indicate in the *Notes* column that the template is PCR product.)

Primer Name: Indicate here the name of the primer to be used for sequencing. (The Core Facility provides at no charge common primers such as T7, T3, SP6, etc. - see full list of available primers at http://www.hmc.psu.edu/core/Genetics_Core/primers.htm.)

If you are using your own primer(s), the Tm should be between 55 and 60°C; thus, the length should be approximately 18-25 nucleotides. (the most accurate T_m estimations are made using "nearest neighbor" calculations, e.g., at http://alces.med.umn.edu/rawtm.html; Primer3 at http://www-genome.wi.mit.edu/cgi-bin/primer/primer3_www.cgi; with our GCG programs; or with the commercial program Oligo.

Notes: should be used for three purposes: 1) you may write your own notes for reference here, 2) request any special reaction conditions required eg. DMSO to minimize 2° structure in GC rich templates, 3) indicate if the DNA template is anything other than plasmid DNA, eg. BAC, PCR product, etc.(for BAC or PAC samples provide either primer Tm or actual sequence for Tm determination).